

***SYSTEMATIC LITERATURE REVIEW : MEDIA KULTUR
JARINGAN TANAMAN DALAM PRODUKSI KUERSETIN***

SKRIPSI

**GRACE IVANA CLARISSA
A 242 041**



**SEKOLAH TINGGI FARMASI INDONESIA
YAYASAN HAZANAH
BANDUNG
2025**

***SYSTEMATIC LITERATURE REVIEW : MEDIA KULTUR
JARINGAN TANAMAN DALAM PRODUKSI KUERSETIN***

SKRIPSI

Sebagai salah satu syarat untuk memperoleh gelar Sarjana Farmasi

**GRACE IVANA CLARISSA
A 242 041**



**SEKOLAH TINGGI FARMASI INDONESIA
YAYASAN HAZANAH
BANDUNG
2025**

***SYSTEMATIC LITERATURE REVIEW : MEDIA KULTUR
JARINGAN TANAMAN DALAM PRODUKSI KUERSETIN***

**GRACE IVANA CLARISSA
A 242 041**

Oktober 2025

Disetujui oleh :

Pembimbing

A handwritten signature in blue ink, consisting of a large loop followed by several vertical strokes and a final flourish.

Himalaya Wana Kelana, M.Pd

Pembimbing

A handwritten signature in blue ink, starting with a large 'S' and followed by 'y' and 'i' with a horizontal stroke.

Dr. Syarif Hamdani, M.Si

Kutipan atau saduran baik sebagian ataupun seluruh naskah, harus menyebut nama pengarang dan sumber aslinya, yaitu Sekolah Tinggi Farmasi Indonesia.

“Sebab Tuhan, Dia sendiri akan berjalan di depanmu, Dia sendiri akan menyertai engkau, Dia tidak akan membiarkan engkau dan tidak akan meninggalkan engkau; janganlah takut dan janganlah patah hati.”

Ulangan 31: 8

SYSTEMATIC LITERATURE REVIEW : MEDIA KULTUR JARINGAN TANAMAN DALAM PRODUKSI KUERSETIN

ABSTRAK

Kuersetin adalah flavonoid dengan aktivitas antioksidan, antiinflamasi, dan antikanker, namun produksinya secara konvensional terbatas oleh faktor lingkungan. Kultur jaringan tanaman menjadi solusi untuk meningkatkan ketersediaan metabolit sekunder secara terkontrol dan berkelanjutan. Komposisi media kultur merupakan faktor kunci yang memengaruhi pertumbuhan jaringan maupun akumulasi metabolit sekunder. Kajian ini merupakan *Systematic Literature Review* berbasis PICOC menggunakan data dari ProQuest, PubMed, ScienceDirect, dan ResearchGate dari tahun 2014–2025. Data dianalisis dengan alur PRISMA untuk menentukan kombinasi media, ZPT, dan elisitor yang paling umum digunakan serta komposisi media menurut metode kultur dan jenis eksplan. Tiga puluh (30) studi yang ditelaah, media Murashige–Skoog (MS) mendominasi 90% penelitian, diikuti Gamborg B5 (6,67%) dan Schenk & Hildebrandt (3,33%). Kultur kalus menggunakan 2,4-D dan BAP menghasilkan produktivitas tertinggi 419,76 mg/g DW; kultur suspensi sel dengan NAA dan Kinetin mencapai 209,93 mg/g DW; kultur organ dengan biostimulator alami mencapai 64,9 mg/g DW; dan kultur akar rambut hasil transformasi *Agrobacterium* tanpa ZPT eksogen menghasilkan 48,24 mg/g DW. Penambahan elisitor seperti metil jasmonat (MeJA), UV-B, dan cahaya biru terbukti meningkatkan biosintesis kuersetin. Eksplan daun paling efektif untuk kultur kalus dan akar rambut, sedangkan nodus untuk kultur suspensi sel dan organ. Optimalisasi media memerlukan pendekatan spesifik berdasarkan jenis tanaman, metode kultur, dan eksplan.

Kata kunci : Kuersetin, Kultur jaringan, Media tumbuh, Eksplan, PICOC

SYSTEMATIC LITERATURE REVIEW: PLANT CULTURE IN VITRO MEDIUM FOR QUERCETIN PRODUCTION

ABSTRACT

Quercetin is a flavonoid with antioxidant, anti-inflammatory, and anticancer activities; however, its conventional production is limited by environmental factors. Plant tissue culture provides a controlled and sustainable approach to enhance the availability of secondary metabolites. The composition of the culture medium is a key factor influencing tissue growth and secondary metabolite accumulation. This study is a PICOC-based Systematic Literature Review using data from ProQuest, PubMed, ScienceDirect, and ResearchGate (2014–2025), analyzed using the PRISMA framework to determine the most common combinations of media, plant growth regulators (PGRs), and elicitors, as well as medium composition according to culture methods and explant types. Among thirty (30) reviewed studies, Murashige–Skoog (MS) medium dominated 90% of applications, followed by Gamborg (B5) (6.67%) and Schenk & Hildebrandt (SH) (3.33%). Callus cultures using 2,4-D + BAP achieved the highest productivity (419.76 mg/g DW); cell suspensions with NAA + Kinetin reached 209.93 mg/g DW; organ cultures with natural biostimulators produced 64.9 mg/g DW; and Agrobacterium-induced hairy roots without exogenous PGRs produced 48.24 mg/g DW. The addition of elicitors such as methyl jasmonate, UV-B, and blue light was found to enhance quercetin biosynthesis. Leaf explants were most effective for callus and hairy roots, while nodes supported cell and organ cultures. Optimization requires species-, culture-, and explant-specific approaches.

Keywords: *Quercetin, Tissue culture, Growth medium, Explants, PICOC*

KATA PENGANTAR

Puji syukur ke hadirat Tuhan Yang Maha Esa atas berkat kebaikan dan kemurahan-Nya sehingga peneliti dapat menyelesaikan Tugas Akhir yang berjudul **“Systematic Literature Review : Media Kultur Jaringan Tanaman dalam Produksi Kuersetin”**. Tugas Akhir ini merupakan syarat wajib bagi setiap mahasiswa agar dapat menyelesaikan.

Dalam penyusunan Tugas Akhir ini, peneliti banyak mendapat bimbingan dan arahan dari berbagai pihak sehingga pada kesempatan ini peneliti ingin mengucapkan terimakasih kepada :

1. Tuhan Yesus yang selalu memberi peneliti hikmat, kekuatan, kesehatan, kelancaran, dan kebahagiaan untuk menyelesaikan Tugas Akhir ini,
2. Dr. Apt. Adang Firmansyah, M.Si. selaku Ketua Sekolah Tinggi Farmasi Indonesia,
3. Dr. apt. Diki Prayugo, M.Si., selaku Wakil Ketua I Bidang Akademik,
4. Dr. Apt. Hesti Riasari, M.Si. selaku Ketua Program Studi Sarjana Farmasi Sekolah Tinggi Farmasi Indonesia
5. Himalaya Wana Kelana, M.Pd. selaku Dosen Pembimbing Utama yang telah membimbing dan memberikan petunjuk serta saran-saran selama penelitian hingga selesainya Tugas Akhir ini.
6. Bapak Dr. Syarif Hamdani, M.Si selaku Dosen Pembimbing Serta yang juga telah banyak membimbing dan memberi masukan selama penelitian hingga selesainya Tugas Akhir ini.
7. Teristimewa kepada Poltak Palmer Marojahan Saragi Napitu, S.H., M.Si dan Monny Simanjuntak, S.Pd selaku kedua orang tua penulis, yang selalu mendoakan, memberi dukungan moril maupun materil, dan memotivasi agar peneliti dapat menyelesaikan tugas akhir ini. Terima kasih untuk semua cinta dan kasih sayang yang diberikan kepada penulis sampai akhirnya skripsi ini rampung.
8. Saudara/i terkasih, Elizabeth Laura Napitu dan Samuel Gicele Napitu yang senantiasa memberi semangat dan menjadi tempat bagi peneliti berkeluh kesah, serta selalu menemani peneliti dalam keadaan apapun.
9. Siti Rahma Dinta, Anzalna Rachman, dan keluarga Literasi Minimal, yang telah kebersamai perjalanan ini dan memberikan banyak warna selama masa perkuliahan hingga penyusunan skripsi.
10. Peneliti sendiri. Terima kasih kepada diri sendiri yang telah berjuang dan bertahan sejauh ini. Terima kasih sudah memilih untuk terus melangkah, meski tidak semua jalan terasa mudah. Terima kasih telah percaya pada proses, pada waktu, dan pada rancangan Tuhan yang selalu lebih indah daripada apa pun yang pernah direncanakan.

Dalam penyusunan skripsi ini masih banyak kesalahan dan kekurangan karena pengetahuan yang masih sangat terbatas. Oleh karena itu, dengan kerendahan hati diharapkan masukan berupa kritik dan saran yang bersifat membangun untuk perbaikan di masa yang akan datang. Penulis berharap semoga tugas akhir ini akan memberikan manfaat bagi penulis sendiri dan juga bagi pihak lain yang berkepentingan.

Bandung, 1 Oktober 2025
Penulis

DAFTAR ISI

LEMBAR PENGESAHAN.....	i
KUTIPAN	ii
PERSEMBAHAN	iii
ABSTRAK	iv
<i>ABSTRACT</i>	v
KATA PENGANTAR.....	vi
DAFTAR ISI.....	viii
DAFTAR TABEL.....	ix
DAFTAR GAMBAR.....	x
DAFTAR LAMPIRAN.....	xi
BAB I PENDAHULUAN	1
1.1 Latar Belakang	1
1.2 Identifikasi Masalah	2
1.3 Tujuan Penelitian.....	2
1.4 Kegunaan Penelitian.....	2
1.5 Waktu dan Tempat Penelitian	2
BAB II TINJAUAN PUSTAKA	3
2.1 Kultur Jaringan	3
2.2 Kuersetin	13
BAB III TATA KERJA	17
3.1 Alat	17
3.2 Bahan.....	17
3.3 Metode Penelitian.....	17
3.4 Metode Pengumpulan Data	19
3.5 Metode Analisis Data	20
3.6 Publikasi	20
BAB IV HASIL PENELITIAN DAN PEMBAHASAN	21
BAB V SIMPULAN DAN ALUR PENELITIAN SELANJUTNYA	31
DAFTAR PUSTAKA	32

DAFTAR TABEL

Tabel 2.1 Komposisi Media MS, B5, WPM, SH.....	10
Tabel 4.1 Media Kultur Jaringan.....	22

DAFTAR GAMBAR

Gambar 2.1 Rumus Struktur Kuersetin	13
Gambar 2.1 Jalur Biosintesis Kuersetin	14
Gambar 3.1 Diagram Alir PRISMA	19
Gambar 4.1 Pemetaan Hasil Produksi Kuersetin pada Berbagai Sistem Kultur <i>In Vitro</i>	24

DAFTAR LAMPIRAN

Lampiran	Halaman
1. <i>Letter of Acceptance</i>	39
2. Draft Jurnal.....	40

DAFTAR PUSTAKA

- Ahmadpoor, F., Zare, N., Asghari, R., & Sheikhzadeh, P. (2022). Sterilization protocols and the effect of plant growth regulators on callus induction and secondary metabolites production in in vitro cultures *Melia azedarach* L. *AMB Express*, 12(1). <https://doi.org/10.1186/s13568-022-01343-8>
- Al-Khayri, J. M., & Naik, P. M. (2020). Elicitor-induced production of biomass and pharmaceutical phenolic compounds in cell suspension culture of date palm (*Phoenix dactylifera* L.). *Molecules*, 25(20). <https://doi.org/10.3390/molecules25204669>
- Alenizi, A., Shibli, R. A. A., Tahtamouni, R. W., Al-Qudah, T. S., & Remeilh, B. A. (2020). In vitro propagation and enhancement of quercetins and isorhamnetin production in wild *paronychia argentea* L. *Jordan Journal of Pharmaceutical Sciences*, 13(1), 65–75.
- Armita, D. (2020). Pemuliaan tanaman melalui fusi protoplas. *Jurnal Biologi Universitas Andalas*, 8(2), 42–47.
- Asra, R., Samarlina, R. A., & Silalahi, M. (2020). Hormon Tumbuhan. In *UKI Press* (Vol. 53, Issue 9).
- Balasubramanian, M., & Girija, S. (2024). Overexpression of AtMYB12 transcription factor simultaneously enhances quercetin-dependent metabolites in radish callus. *Heliyon*, 10(8), e27053. <https://doi.org/10.1016/j.heliyon.2024.e27053>
- Balasubramanian, M., Kumaraguru, G., & Girija, S. (2024). Efficient extraction methods and callus culture of quercetin from Indian white radish: yield analysis via HPLC quantification and assessment of cytotoxic activity. *International Journal of Food Science and Technology*, 59(6), 3556–3572. <https://doi.org/10.1111/ijfs.17043>
- Bandekar, H., & Lele, S. S. (2014). Production of flavonol Quercetin from cultured plant cells of Banyan (*Ficus benghalensis* L .). *International Journal of Innovative Research in Science, Engineering and Technology*, 3(5), 12150–12157.
- Barbulova, A., Apone, F., & Colucci, G. (2014). Plant cell cultures as source of cosmetic active ingredients. *Cosmetics*, 1(2), 94–104. <https://doi.org/10.3390/cosmetics1020094>
- Barus, E. M., & Restuati, M. (2018). Pengaruh Media Kultur pada Planlet Kentang *Solanum tuberosum* L terhadap Totipensi Pertumbuhan Tunas. *Jurnal Ilmiah Farmasi Imelda*, 1(2), 51–56. <https://jurnal.uimedan.ac.id/index.php/Jurnalfarmasi51Journalhomepage:https://jurnal.uimedan.ac.id/index.php/Jurnalfarmasi>
- Biswas, D., Chakraborty, A., Mukherjee, S., & Ghosh, B. (2023). Hairy root culture: a potent method for improved secondary metabolite production of Solanaceous plants. *Frontiers in Plant Science*, 14(September), 1–19. <https://doi.org/10.3389/fpls.2023.1197555>
- Cardoso, J. C., de Oliveira, M. E. B. S., & Cardoso, F. de C. I. (2019). Advances and challenges on the in vitro production of secondary metabolites from medicinal plants. *Horticultura Brasileira*, 37(2), 124–132. <https://doi.org/10.1590/s0102-053620190201>
- Chimdessa, E., & Agricultural, O. (2020). Composition and Preparation of Plant

- Tissue Culture Medium. *Journal of Tissue Culture and Bioengineering*, 3(01).
<https://doi.org/10.29011/2688-6502.000020>
- Coskun, Y., & Yaman, V. (2022). Polyamine Elicitation of Quercetin and Rutin Production in Callus Cultures of Caper and Impact to Regeneration. *Journal of Scientific and Industrial Research*, 81(5), 475–481.
<https://doi.org/10.56042/jsir.v81i05.58826>
- Dar, S. A., Nawchoo, I. A., Tyub, S., & Kamili, A. N. (2021). Effect of plant growth regulators on in vitro induction and maintenance of callus from leaf and root explants of *Atropa acuminata* Royal ex Lindl. *Biotechnology Reports*, 32(August), e00688. <https://doi.org/10.1016/j.btre.2021.e00688>
- Dr. Ir. Dwi Hapsoro, M. S., & Prof. Dr. Ir. Yusnita, M. S. (2020). *KULTUR JARINGAN - Teori dan Praktik*. Andi Publisher.
<https://books.google.co.id/books?id=PyOwDwAAQBAJ>
- Durán, M. D. L., Zabala, M. E. A., & Londoño, G. A. C. (2021). Optimization of Flavonoid Production in Plant Cell Culture of *Thevetia peruviana* Elicited with Methyl Jasmonate and Salicylic Acid. *Brazilian Archives of Biology and Technology*, 64, 1–12. <https://doi.org/10.1590/1678-4324-2021210022>
- Ferreira, F. M. L., Rius, S. P., & Casati, P. (2012). Flavonoids: Biosynthesis, biological functions, and biotechnological applications. *Frontiers in Plant Science*, 3(SEP), 1–15. <https://doi.org/10.3389/fpls.2012.00222>
- Gabr, A. M. M., Fayek, N. M., Mahmoud, H. M., El-Bahr, M. K., Ebrahim, H. S., Sytar, O., & El-Halawany, A. M. (2022). Effect of Light Quality and Media Components on Shoot Growth, Rutin, and Quercetin Production from Common Buckwheat. *ACS Omega*, 7(30), 26566–26572.
<https://doi.org/10.1021/acsomega.2c02728>
- George, E., Hall, M., & De Klerk, J. (1984). Plant Propagation by Tissue Culture 3rd Edition Volume 1 The Background. In *Springer*.
- Guru, A., Dwivedi, P., Kaur, P., & Pandey, D. K. (2022). Exploring the role of elicitors in enhancing medicinal values of plants under in vitro condition. *South African Journal of Botany*, 149, 1029–1043.
<https://doi.org/10.1016/j.sajb.2021.10.014>
- Halder, M., Sarkar, S., & Jha, S. (2019). Elicitation: A biotechnological tool for enhanced production of secondary metabolites in hairy root cultures. *Engineering in Life Sciences*, 19(12), 880–895.
<https://doi.org/10.1002/elsc.201900058>
- Handoko, R. N. S., & Lin, S. Y. (2025). Integrating plant growth regulators and biostimulants to enhance resilient and sustainable raspberry and blackberry production. *Scientia Horticulturae*, 350(July), 114296.
<https://doi.org/10.1016/j.scienta.2025.114296>
- Herawan, T., & Leksono, B. (2018). Regenerasi in vitro *Eucalyptus pellita* F. Muell menggunakan kultur mata tunas. *Jurnal Perbenihan Tanaman Hutan*, 6(1), 1–13.
- Huang, X., Yao, J., Zhao, Y., Xie, D., Jiang, X., & Xu, Z. (2016). Efficient rutin and quercetin biosynthesis through flavonoids-related gene expression in *Fagopyrum tataricum* gaertn. Hairy root cultures with UV-B irradiation. *Frontiers in Plant Science*, 7(FEB2016), 1–11.
<https://doi.org/10.3389/fpls.2016.00063>
- Humbal, A., & Pathak, B. (2023). Influence of exogenous elicitors on the

- production of secondary metabolite in plants: A review ('“VSI: secondary metabolites”’). *Plant Stress*, 8(May), 100166. <https://doi.org/10.1016/j.stress.2023.100166>
- Hussain, M. J., Abbas, Y., Nazli, N., Fatima, S., Drouet, S., Hano, C., & Abbasi, B. H. (2022). Root Cultures, a Boon for the Production of Valuable Compounds: A Comparative Review. *Plants*, 11(3). <https://doi.org/10.3390/plants11030439>
- Iqbal, M. S., Iqbal, Z., & Ansari, M. I. (2019). Enhancement of total antioxidants and flavonoid (quercetin) by methyl jasmonate elicitation in tissue cultures of onion (*Allium cepa* L.). *Acta Agrobotanica*, 72(3), 1–11. <https://doi.org/10.5586/aa.1784>
- Istiqomah, M., Setiari, N., & Nurchayati, Y. (2020). Pengaruh Media MS dan VW Terhadap Pertumbuhan Planlet Anggrek Bulan Setelah Transplanting. *Prosiding Seminar Nasional Pendidikan Biologi Dan Saintek Ke-5, Semarang*, 476–480.
- Karakas, F. P. (2020). Efficient plant regeneration and callus induction from nodal and hypocotyl explants of goji berry (*Lycium barbarum* L.) and comparison of phenolic profiles in calli formed under different combinations of plant growth regulators. *Plant Physiology and Biochemistry*, 146, 384–391. <https://doi.org/10.1016/j.plaphy.2019.11.009>
- Karti, P. D. M. H., Wijayanti, I., & Pramadi, S. D. (2020). Teknik Aklimatisasi Pada Tanaman Lamtoro (*Leucaena Leucocephala*) dengan Perbedaan Media Tanam dan Sifat Tumbuh. *Pastura*, 10(1), 46. <https://doi.org/10.24843/pastura.2020.v10.i01.p11>
- Karuppaiya, P., & Tsay, H. S. (2020). Enhanced production of podophyllotoxin, kaempferol, and quercetin from callus culture of *Dioscorea pleiantha* (Hance) Woodson: An endangered medicinal plant. *Biotechnology and Applied Biochemistry*, 67(1), 95–104. <https://doi.org/10.1002/bab.1810>
- Khajeh, H., Fazeli-Nasab, B., Salehi Sardoei, A., Fotoohiyan, Z., Hatami, M., Mirzaei, A., Ghorbanpour, M., & Maggi, F. (2025). Characterization of Biological Components of Leaves and Flowers in *Moringa peregrina* and Their Effect on Proliferation of *Staurogyne repens* in Tissue Culture Conditions. *Plants*, 14(15), 2340. <https://doi.org/10.3390/plants14152340>
- Khoirunnisa, I., & Sumiwi, S. A. (2019). Review Artikel: Peran Flavonoid Pada Berbagai Aktivitas Farmakologi. *Farmaka*, 17(2), 131–142. <https://jurnal.unpad.ac.id/farmaka/article/view/21922>
- Krishnan, R., Anil Kumar, V. S., & Murugan, K. (2014). Establishment of cell suspension culture in *Marchantia linearis* Lehm & Lindenb. For the optimum production of flavonoids. 3 *Biotech*, 4(1), 49–56. <https://doi.org/10.1007/s13205-013-0123-7>
- Kurnia, A., Rahma, D., Fadhilah, H., Sari, M., Putri, P. A., & Advinda, L. (2022). Effect of IAA and BAP Differences on Patchouli Plant Growth (*Pogostemon cablin* Benth) In-Vitro. *Prosiding SEMNAS BIO*, 2(2), 758–765.
- Larsen, D. H., Li, H., Shrestha, S., Verdonk, J. C., Nicole, C. C. S., Marcelis, L. F. M., & Woltering, E. J. (2022). Lack of Blue Light Regulation of Antioxidants and Chilling Tolerance in Basil. *Frontiers in Plant Science*, 13(April), 1–16. <https://doi.org/10.3389/fpls.2022.852654>
- Li, Y., Yao, J., Han, C., Yang, J., Chaudhry, M. T., Wang, S., Liu, H., & Yin, Y.

- (2016). Quercetin, inflammation and immunity. *Nutrients*, 8(3), 1–14. <https://doi.org/10.3390/nu8030167>
- Long, Y., Yang, Y., Pan, G., & Shen, Y. (2022). New Insights Into Tissue Culture Plant-Regeneration Mechanisms. *Frontiers in Plant Science*, 13(June). <https://doi.org/10.3389/fpls.2022.926752>
- Madanakumar, A. J., Murukan, G., Lawarence, B., & Kumaraswamy, M. (2017). Isolation, purification of quercetin from in vitro cell suspension culture of caesalpinia pulcherrima and its analysis by HPLC-DAD and NMR. *Pharmacognosy Journal*, 9(6), s44–s51. <https://doi.org/10.5530/pj.2017.6s.156>
- Mahadi, I., Syafi'i, W., & Sari, Y. (2016). Callus Induction of Calamansi (Citrus microcarpa) Using 2,4-D and BAP Hormones by in vitro Methods. *Jurnal Ilmu Pertanian Indonesia*, 21(2), 84–89. <https://doi.org/10.18343/jipi.21.2.84>
- Matvieieva, N., Bessarabov, V., Khainakova, O., Duplij, V., Bohdanovych, T., Ratushnyak, Y., Kuzmina, G., Lisovyi, V., Zderko, N., & Kobylinska, N. (2023). Cichorium intybus L. “hairy” roots as a rich source of antioxidants and anti-inflammatory compounds. *Heliyon*, 9(3), e14516. <https://doi.org/10.1016/j.heliyon.2023.e14516>
- Maulida, D., Erfa, L., & Negeri Lampung, P. (2020). Kultur Embrio Kelapa Kopyor Menggunakan Beberapa Konsentrasi BA dan Air Kelapa Kopyor Coconut Embrio Culture Uses Some Concentrations of BA and Coconut Water. *Jurnal Penelitian Pertanian Terapan*, 20(3), 247–251. <http://www.jurnal.polinela.ac.id/JPPTeISSN2047-1781>
- Meresa, B. K., Ayimut, K. M., Weldemichael, M. Y., Geberemedhin, K. H., Kassegn, H. H., Geberemikael, B. A., & Egigu, E. M. (2024). Carbohydrate elicitor-induced plant immunity: Advances and prospects. *Heliyon*, 10(15), e34871. <https://doi.org/10.1016/j.heliyon.2024.e34871>
- Motolinia-Alcántara, E. A., Franco-Vásquez, A. M., Nieto-Camacho, A., Arreguín-Espinosa, R., Rodríguez-Monroy, M., Cruz-Sosa, F., & Román-Guerrero, A. (2023). Phenolic Compounds from Wild Plant and In Vitro Cultures of Ageratina pichichensis and Evaluation of Their Antioxidant Activity. *Plants*, 12(5). <https://doi.org/10.3390/plants12051107>
- Mounika, K., & Giri, A. (2024). Elicitation an approach to improving the metabolic profile of therapeutically important flavonoids from callus cultures of Alpinia purpurata. *Agricultural and Biological Research*, 40(1), 792–800. <https://doi.org/10.35248/0970-1907.24.40.792-800>
- Najhah, M. Y., Jaafar, H. Z. E., Nakasha, J. J., & Hakiman, M. (2021). Shoot Multiplication and Callus Induction of Labisia pumila var. alata as Influenced by Different Plant Growth Regulators Treatments and Its Polyphenolic Activities Compared with the Wild Plant. *Molecules*, 26(3229), 1–14.
- Nasution, T. R., Agroteknologi, P. S., Pertanian, F., Peternakan, D. A. N., Islam, U., Sultan, N., & Kasim, S. (2018). Pengaruh Konsentrasi Bap Dan Naa Terhadap Induksi Kalus Daun Pasakbumi (Eurycoma Longifoliajack) Secara In Vitro. Riau: Universitas Islam Negeri Sultan Syarif Kasim Riau.
- Naushafira, N. D., Hanuna, H., Dewi, M. K., & Selius, W. V. N. (2022). Kajian Sistematis: Aktivitas Kuersetin sebagai Inhibitor Kanker Payudara secara In Vitro. *Generics: Journal of Research in Pharmacy*, 2(2), 105–112. <https://doi.org/10.14710/genres.v2i2.15774>

- Ndruru, H. S., Telambanua, P. H., Nazara, R. V., & Gulo, S. D. (2024). Pemanfaatan Elisitor pada Tanaman. *Sapta Agrica*, 3(1), 39–51.
- Negri, S., Gambini, S., Ceoldo, S., Avesani, L., Commisso, M., & Guzzo, F. (2021). Undifferentiated in vitro cultured actinidia deliciosa as cell factory for the production of quercetin glycosides. *Plants*, 10(11). <https://doi.org/10.3390/plants10112499>
- Ozyigit, I. I., Dogan, I., Hocaoglu-Ozyigit, A., Yalcin, B., Erdogan, A., Yalcin, I. E., Cabi, E., & Kaya, Y. (2023). Production of secondary metabolites using tissue culture-based biotechnological applications. *Frontiers in Plant Science*, 14(June), 1–28. <https://doi.org/10.3389/fpls.2023.1132555>
- Popova, P., Zarev, Y., & Ionkova, I. (2021). Biotransformation of quercetin, kaempferol and apigenin to monoglycosylated derivatives by in vitro suspension cultures of *Astragalus vesicarius* ssp. *carniolicus*. *Pharmacia*, 68(2), 307–311. <https://doi.org/10.3897/Pharmacia.68.E54289>
- Ramirez-Estrada, K., Vidal-Limon, H., Hidalgo, D., Moyano, E., Golenioswki, M., Cusidó, R. M., & Palazon, J. (2016). Elicitation, an effective strategy for the biotechnological production of bioactive high-added value compounds in plant cell factories. *Molecules*, 21(2). <https://doi.org/10.3390/molecules21020182>
- Rebaz, N., Dalar Van Yuzuncu, A., Ahmet Ozdemir, F., & Turker Van Yuzuncu, M. (2019). In vitro propagation and secondary metabolites investigation of *hypericum perforatum* L. *Fresenius Environmental Bulletin*, 28(7), 5569–5576. <https://www.researchgate.net/publication/334049867>
- Robles-Martínez, M., Barba-De la Rosa, A. P., Guéraud, F., Negre-Salvayre, A., Rossignol, M., & Santos-Díaz, M. D. S. (2016). Establishment of callus and cell suspensions of wild and domesticated *Opuntia* species: study on their potential as a source of metabolite production. *Plant Cell, Tissue and Organ Culture*, 124(1), 181–189. <https://doi.org/10.1007/s11240-015-0886-0>
- Roostika, I., Sutanto, A., Edison, N., & Dewi, N. (2019). Kultur Embrio Pisang Liar *Musa acuminata* ssp. *sumatrana* yang Langka (Embryo Culture of Endangered Wild Banana *Musa acuminata* ssp. *sumatrana*). *Jurnal Hortikultura*, 28(1), 25. <https://doi.org/10.21082/jhort.v28n1.2018.p25-32>
- Sajjalaguddam, R. R., & Paladugu, A. (2015). Phenylalanine enhances quercetin content in in vitro cultures of *abutilon indicum* L. *Journal of Applied Pharmaceutical Science*, 5(10), 80–84. <https://doi.org/10.7324/JAPS.2015.501014>
- Septiawati, N., Dalimunthe, A., Hasibuan, S., & Aziz, R. (2021). Penggunaan Air Kelapa dan Indol-3-Butyric-Acid Iba Untuk Induksi Multiplikasi Tunas Eksplan Tanaman Kentang (*Solanum Tuberosum* L.) Secara In-Vitro Cocconut Water and Indol-3-Butyric-Acid (IBA) for Induction of Shot Multiplication from Potato (*Solanum Tube*. *Jurnal Ilmiah Pertanian (JIPERTA)*, 3(1), 76–85. <http://jurnalmahasiswa.uma.ac.id/index.php/jiperta>
- Seran, S., Boobalan, S., Keerthanaa, T., Kumaravel, S., Karunakaran, G., & Srinivasan, R. (2025). Augmentation of betacyanin and quercetin in hybrid callus: Comprehensive assessment of biosynthesized silver nanoparticles for their potent biological activities, advanced in silico interactions, and rigorous toxicological evaluation. *Industrial Crops and Products*, 227(March), 120824. <https://doi.org/10.1016/j.indcrop.2025.120824>
- Setiawati, T., Arofah, A. N., Nurzaman, M., Annisa, A., Mutaqin, A. Z., & Hasan,

- R. (2023). Effect of sucrose as an elicitor in increasing quercetin-3-O-rhamnoside (quercitrin) content of chrysanthemum (*Chrysanthemum morifolium* Ramat) callus culture based on harvest time differences. *Biotechnologia*, 104(3), 289–300. <https://doi.org/10.5114/bta.2023.130731>
- Setiawati, T., Ayalla, A., Nurzaman, M., Kusumaningtyas, V. A., & Bari, I. (2020). Analysis of Secondary Metabolites of Shoot, Callus Culture and Field Plant of *Chrysanthemum morifolium* Ramat. *Jurnal ILMU DASAR*, 21(1), 1. <https://doi.org/10.19184/jid.v21i1.8665>
- Setiawati, T., Zazuli, S. F., Annisa, A., Nurzaman, M., & Irawan, B. (2020). Pengaruh Polyethylene Glycol (Peg) Terhadap Kadar Kuersetin Kultur Kalus *Chrysanthemum Morifolium* Ramat pada Kondisi Pencahayaan Berbeda. *Al-Kauniyah: Jurnal Biologi*, 13(1), 116–127. <https://doi.org/10.15408/kauniyah.v13i1.13688>
- Sidik, N. J., Agha, H. M., Alkamil, A. A., Alsayadi, M. M. S., & Mohammed, A. A. (2024). A Mini Review of Plant Tissue Culture: The Role of Media Optimization, Growth Regulators in Modern Agriculture, Callus Induction and the Applications. *AUIQ Complementary Biological System*, 1(2), 96–109. <https://doi.org/10.70176/3007-973x.1019>
- Sinaulan, J. S., Lengkong, E. F., & Tulung, S. (2019). Respon Pembentukan Kalus Embriolik Tanaman Krisan Kulo (*Chysanthemum morifolium*) terhadap Pemberian Zat Pengatur Tumbuh Sitokinin. *Jurnal Unsrat*, 1(2), 1–4.
- Syamsinar. (2022). Efektivitas Media B5 (GAMBORG) untuk Multiplikasi Sengon (*Paraserianthes falcataria* L. Nielsen) secara In Vitro [Universitas Hasanuddin]. In *Skripsi* (Vol. 1). https://repository.unhas.ac.id/id/eprint/18772/2/M011181006_skripsi_01-09-2022_1-2.pdf
- Taji, A., Kumar, P., & Lakshmanan, P. (2024). In Vitro Plant Breeding. *In Vitro Plant Breeding*, 1–167. <https://doi.org/10.1201/9781003578529>
- Thwe, A., Arasu, M. V., Li, X., Park, C. H., Kim, S. J., Al-Dhabi, N. A., & Park, S. U. (2016). Effect of different *Agrobacterium rhizogenes* strains on hairy root induction and phenylpropanoid biosynthesis in tartary buckwheat (*Fagopyrum tataricum* Gaertn). *Frontiers in Microbiology*, 7(MAR), 1–10. <https://doi.org/10.3389/fmicb.2016.00318>
- Tossi, V. E., Regalado, J. J., Iannicelli, J., Laino, L. E., Burrieza, H. P., Escandón, A. S., & Pitta-Álvarez, S. I. (2019). Beyond arabidopsis: Differential UV-B response mediated by UVR8 in diverse species. *Frontiers in Plant Science*, 10(June), 1–17. <https://doi.org/10.3389/fpls.2019.00780>
- Tuwo, M., Tambaru, E., & Marianty, N. (2022). Respon Pertumbuhan Biji Jeruk Keprok *Citrus reticulata* Blanco pada Beberapa Teknik Sterilisasi. *J Ilmu Alam Dan Lingkungan*, 13(2), 32–39.
- Vesnina, A., Milentyeva, I., Minina, V., Kozlova, O., & Asyakina, L. (2023). Evaluation of the In Vivo Anti-Atherosclerotic Activity of Quercetin Isolated from the Hairy Roots of *Hedysarum neglectum* Ledeb. *Life*, 13(8). <https://doi.org/10.3390/life13081706>
- Vollmannová, A., Bojnanská, T., Musilová, J., Lidiková, J., & Cifrová, M. (2024). Quercetin as one of the most abundant represented biological valuable plant components with remarkable chemoprotective effects - A review. *Heliyon*, 10(12). <https://doi.org/10.1016/j.heliyon.2024.e33342>

- Wulandari, S., Nisa, Y. S., Taryono, T., Indarti, S., & Sayekti, R. S. (2022). Sterilisasi Peralatan dan Media Kultur Jaringan. *Agrotechnology Innovation (Agrinova)*, 4(2), 16. <https://doi.org/10.22146/a.77010>
- Ying, W., Wen, G., Xu, W., Liu, H., Ding, W., Zheng, L., He, Y., Yuan, H., Yan, D., Cui, F., Huang, J., Zheng, B., & Wang, X. (2023). *Agrobacterium rhizogenes*: paving the road to research and breeding for woody plants. *Frontiers in Plant Science*, 14 (November). <https://doi.org/10.3389/fpls.2023.1196561>
- Yuniardi, F. (2020). Aplikasi Dimmer Switch pada Rak Kultur Sebagai Pengatur Kebutuhan Intesitas Cahaya Optimum Bagi TanamanIn Vitro. *Indonesian Journal of Laboratory*, 1(4), 8. <https://doi.org/10.22146/ijl.v1i4.52991>
- Zafari, S., Sharifi, M., & Chashmi, N. A. (2018). A comparative study of biotechnological approaches for producing valuable flavonoids in *Prosopis farcta*. *Cytotechnology*, 70(2), 603–614. <https://doi.org/10.1007/s10616-017-0143-y>
- Zhang, X., Ahmad, N., Zhang, Q., Wakeel Umar, A., Wang, N., Zhao, X., Zhou, K., Yao, N., & Liu, X. (2023). Safflower Flavonoid 3'5'Hydroxylase Promotes Methyl Jasmonate-Induced Anthocyanin Accumulation in Transgenic Plants. *Molecules*, 28(7). <https://doi.org/10.3390/molecules28073205>